

**Acknowledgment.** We thank Professor G. C. Dismukes and D. Abramowicz for assistance with the oxygen electrode measurements and Dr. M. Benecky for helpful discussions. This work was supported by NSF Grant CHE 8106084 and NIH Grant GM25158.

**Registry No.** Glucose oxidase, 9001-37-0.

**Pyridinium-1-ylcarbons:  
1,2,3,3-Tetrakis(4-(dimethylamino)pyridinium-1-yl)-  
cyclopropene Tetrachloride and  
1,1,2,3,3-Pentakis(4-(dimethylamino)pyridinium-1-yl)-  
allylide Tetrachloride**

Kenneth C. Waterman and Andrew Streitwieser, Jr.\*

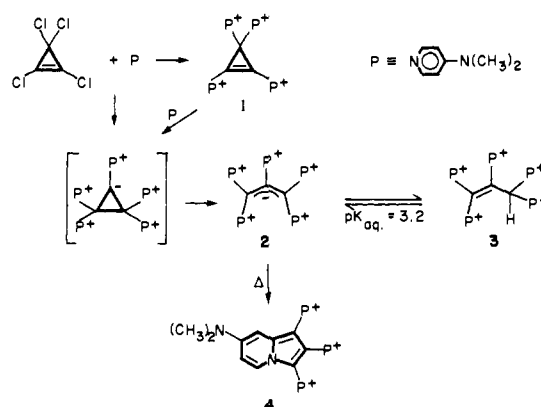
Department of Chemistry, University of California  
Berkeley, California 94720

Received March 29, 1984

Poly-*N*-pyridinium compounds have been studied extensively for their biological activity.<sup>1-15</sup> Compounds having all available positions occupied by pyridinium groups have not been reported in the literature. Such compounds are polyionic, with a high concentration of charge, and may have unusual properties. We report here the synthesis in high yield of two such novel perpyridinium compounds from tetrachlorocyclopropene<sup>16</sup> and 4-(dimethylamino)pyridine (DMAP), 1,2,3,3-tetrakis(4-(dimethylamino)pyridinium-1-yl)cyclopropene tetrachloride (1), and 1,1,2,3,3-pentakis(4-(dimethylamino)pyridinium-1-yl)allylide tetrachloride (2).

Compound 1 was prepared in greater than 60% yield by direct reaction at 0 °C of tetrachlorocyclopropene with a solution of stoichiometric DMAP in 80% methylene chloride and 20% THF. The dry salt appears to be stable but is hygroscopic; the tetrakis(hexafluoroarsenate) is not hygroscopic. In water or methanol solution the compound reacts to give products not yet totally identified. In a chloroform solution, 1 reacts with DMAP to give 2. This reaction supports the proposed intermediacy of 1 in the direct reaction to form 2 (see Scheme I). Identification of 1 is consistent with <sup>1</sup>H NMR,<sup>17</sup> <sup>13</sup>C NMR,<sup>18</sup> and IR (alkene stretch at 1635 cm<sup>-1</sup>) spectroscopy and analysis of the tetrachloride and

**Scheme I**



tetrakis(hexafluoroarsenate) salts.<sup>19</sup>

Compound 2 was prepared in greater than 60% yield by reaction of tetrachlorocyclopropene with DMAP in chloroform. The structure proposed is consistent with high-resolution <sup>1</sup>H NMR,<sup>20</sup> <sup>13</sup>C NMR,<sup>21</sup> and UV-vis ( $\lambda_{\max}$  450 nm) spectroscopy and analysis.<sup>22</sup> This remarkable compound, an allyl anion stabilized by five positive charges, forms dark red crystals that appear to be quite stable in air. Compound 2 can be protonated with HCl or HBF<sub>4</sub> to give the corresponding propene, 3. The chloride salt of 3 shows four distinct pyridiniums in NMR spectroscopy. With the BF<sub>4</sub> salt of 3,<sup>23</sup> exchange is rapid and only three types of pyridiniums are observed by <sup>1</sup>H NMR. Since 2 is intensely colored, and its protonated form 3 is not, the aqueous acidity of 3 could be determined by using the compound itself as the indicator. The pK<sub>a</sub> of 3 is found to be 3.2 ± 0.1. This makes compound 3 the most acidic conjugate acid of an ylide for which data are reported.<sup>24,25</sup>

When 2 was heated to 190 °C under vacuum, 1 equiv of 4-(dimethylamino)pyridine hydrochloride sublimed, leaving 1,2,3-tris(4-(dimethylamino)pyridinium-1-yl)-7-(dimethylamino)indolizine trichloride<sup>26</sup> (4) in greater than 60% yield, consistent with the known reactivity of pyridinium allylides<sup>27,28</sup> (Scheme I). The reaction of pyridine with tetrachlorocyclopropene to form indolizines was reported in an earlier paper.<sup>29</sup> The pyridine reaction apparently involves successive additions to the cyclopropene and elimination of chloride ion; with two and three pyridinium substituents, electrocyclic ring opening of the intermediate cyclopropyl anion competes successfully with further loss of chloride ion. With 4-(dimethylamino)pyridine, however, ring opening does not occur until all of the chlorines have been replaced.

- (1) Corder, C. N.; Way, J. L. *J. Med. Chem.* **1966**, *9*, 638.
- (2) Ashani, Y.; Edery, H.; Zahavy, J.; Kunberg, W.; Cohen, S. *Isr. J. Chem.* **1965**, *3*, 133.
- (3) Luttringhaus, A.; Hagedorn, I. *Arzneim.-Forsch.* **1964**, *14*, 1.
- (4) Ashani, Y.; Cohen, S. *J. Med. Chem.* **1971**, *14*, 621; **1970**, *13*, 471.
- (5) Ashani, Y.; Cohen, S. *Isr. J. Chem.* **1967**, *5*, 59.
- (6) Barfknecht, C. F.; Benz, F. W.; Long, J. P. *J. Med. Chem.* **1971**, *14*, 1003.
- (7) Lamb, J. C.; Steinberg, G. M.; Solomon, S.; Hackley, B. E., Jr. *Biochemistry* **1965**, *4*, 2475.
- (8) Franchetti, P.; Grifantini, M.; Stein, M. L. *J. Pharm. Sci.* **1970**, *59*, 710.
- (9) Barfknecht, C. F.; Long, J. P.; Benz, F. W. *J. Pharm. Sci.* **1970**, *60*, 138.
- (10) Patocka, J. *Collect. Czech. Chem. Commun.* **1971**, *36*, 2677.
- (11) Dirks, E.; Scherer, A.; Schmidt, M.; Zimmer, G. *Arzneim.-Forsch.* **1970**, *20*, 197.
- (12) Schoene, K.; Strake, E. M. *Biochem. Pharmacol.* **1971**, *20*, 1041.
- (13) Kuhnert, H. *Arzneim.-Forsch.* **1970**, *20*, 774.
- (14) Hobbiger, R.; O'Sullivan, D. G.; Sadler, P. W. *Nature (London)* **1958**, *182*, 1498.
- (15) Kewitz, H.; Wilson, I. B.; Nachmansohn, D. *Arch. Biochem. Biophys.* **1956**, *64*, 456.
- (16) Tobey, S. W.; West, R. *J. Am. Chem. Soc.* **1966**, *88*, 2481.
- (17)  $\delta$ , D<sub>2</sub>O (*J* in hertz): 8.21 (4, d, *J* = 7.83), 8.02 (4, d, *J* = 7.81), 7.07 (4, d, *J* = 7.83), 6.91 (4, d, *J* = 7.83), 3.29 (12, s), 3.17 (12, s). CDCl<sub>3</sub>: 9.88 (4, d, *J* = 7.8), 9.47 (4, d, *J* = 7.8), 7.11 (4, d, *J* = 7.8), 6.91 (4, d, *J* = 7.8), 3.35 (12, s), 3.24 (12, s).
- (18)  $\delta$ , D<sub>2</sub>O (*J* in hertz): 163.4 (m), 145.32 (d, *J*<sub>CH</sub> = 189.1), 144.28 (d, *J*<sub>CH</sub> = 185.0), 115.87 (d, *J*<sub>CH</sub> = 174.0), 115.35 (d, *J*<sub>CH</sub> = 172.2), 108.14 (s), 74.77 (s), 47.34 (q, *J*<sub>CH</sub> = 141), 46.69 (q, *J*<sub>CH</sub> = 140).

(19) Analyses by Analytical Services Laboratory, UC College of Chemistry. Anal. Calcd for C<sub>31</sub>H<sub>40</sub>N<sub>5</sub>Cl<sub>4</sub>·6.5H<sub>2</sub>O (chloride): C, 47.5; H, 6.8; N, 14.3; Cl, 18.1. Found: C, 47.3; H, 6.45; N, 14.5; Cl, 18.7. Calcd for C<sub>31</sub>H<sub>40</sub>N<sub>5</sub>As<sub>4</sub>F<sub>24</sub>·(CH<sub>3</sub>COCH<sub>3</sub>) (hexafluoroarsenate): C, 30.5; H, 3.4; N, 8.4. Found: C, 30.4; H, 3.55; N, 8.5; Cl, 0. Acetone was found in the NMR spectrum.

(20)  $\delta$ , D<sub>2</sub>O (*J* in hertz): 7.90 (2, d, *J* = 7.5), 7.59 (4, d, *J* = 7.7), 7.58 (4, d, *J* = 7.6), 6.40 (10, m), 2.81 (12, s), 2.76 (18, s). CD<sub>3</sub>OD: 8.81 (2, d, *J* = 7.66), 8.37 (8, dd, *J* = 7.94, 7.94), 4.93 (10, m), 3.27 (12, s), 3.23 (18, s).

(21)  $\delta$ , D<sub>2</sub>O: 49.6, 50.4, 50.6, 108.9, 117.3, 118.6, 119.1, 138.3, 150.9, 153.0, 153.1, 153.1, 153.5, 165.8, 167.4.

(22) Anal. Calcd for C<sub>38</sub>H<sub>50</sub>N<sub>10</sub>Cl<sub>4</sub>·6H<sub>2</sub>O: C, 50.9; H, 6.9; N, 15.6; Cl, 15.85. Found: C, 51.0; H, 7.1; N, 15.8; Cl, 16.05.

(23) Anal. Calcd for C<sub>38</sub>H<sub>51</sub>N<sub>10</sub>B<sub>3</sub>F<sub>30</sub>·2H<sub>2</sub>O (BF<sub>4</sub> salt): C, 40.8; H, 4.9; N, 12.5. Found: C, 40.7; H, 5.0; N, 12.4; Cl, 0.

(24) Kosower, E. M.; Ramsey, B. G. *J. Am. Chem. Soc.* **1959**, *81*, 856.

(25) Berson, J. A.; Evleth, E. M., Jr.; Hamlet, Z. *J. Am. Chem. Soc.* **1965**, *87*, 2888.

(26) UV-vis (MeOH):  $\lambda_{\max}$  356 nm ( $\log \epsilon$  4.16). <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$  (*J* in hertz): 8.41 (d, 2, *J* = 8.0), 8.38 (2, d, *J* = 8.0), 8.32 (2, d, *J* = 7.8), 7.89 (1, d, *J* = 8.0), 6.95 (1, dd, *J* = 7.9, 2.5), 6.9 (6, m), 6.24 (1, d, *J* = 2.5), 3.36 (6, s), 3.30 (6, s), 3.25 (6, s), 3.04 (6, s). Anal. Calcd for C<sub>31</sub>H<sub>39</sub>N<sub>8</sub>Cl<sub>3</sub>·4.75H<sub>2</sub>O: C, 52.0; H, 6.8; N, 15.7; Cl, 14.9. Found: C, 52.0; H, 6.6; N, 15.5; Cl, 15.2.

(27) Sasaki, T.; Kanematsu, K.; Kakehi, A.; Ito, G. *Tetrahedron* **1972**, *28*, 4947.

(28) Pohjala, E. *Tetrahedron Lett.* **1972**, 2585.

(29) Smith, K. A.; Streitwieser, A., Jr. *J. Org. Chem.* **1983**, *48*, 2629.

There appears to be a subtle and unexpected balance between the rates of cyclopropyl anion ring opening and loss of chloride ion that depends sensitively on the structure of the pyridine.

The first pyridinium-1-ylcarbons, **1** and **2**, show a chemistry that invites further study. They have the potential of opening up a whole new field of organic chemistry in which their high concentration of charge may provide exciting properties.

**Acknowledgment.** This work was supported in part by NSF Grant CHE82-05696.

## Total Synthesis of C<sub>22</sub>-Prostanoids in the E and F Series Based on Docosaehaenoic Acid

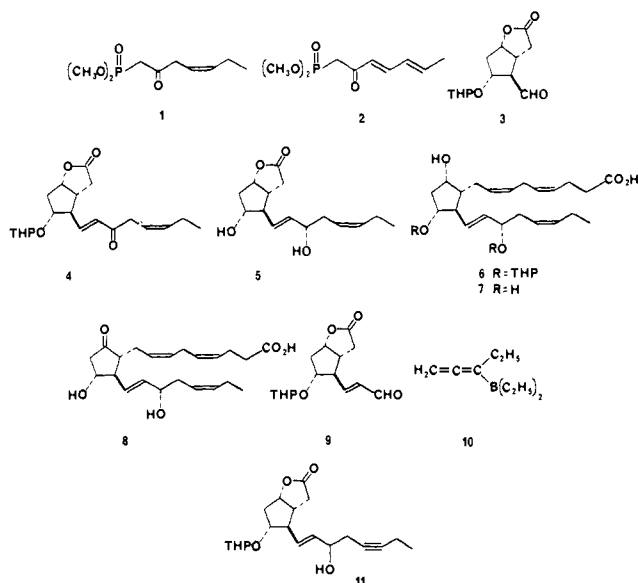
E. J. Corey,\* Shuichi Ohuchida, and Robert Hahl

Department of Chemistry, Harvard University  
Cambridge, Massachusetts 02138

Received March 26, 1984

Docosa-4,7,10,13,16,19-(Z)-hexaenoic acid (docosaehaenoic acid, DCHA) is a major polyunsaturated fatty acid in marine lipid, along with eicosa-5,8,11,14,17-(Z)-pentaenoic acid (EPA). The obvious question as to whether DCHA might be the progenitor of a family of C<sub>22</sub>-prostanoids, analogous to those derived from arachidonic acid or EPA, was made more intriguing by the reported isolation of a C<sub>22</sub>-prostanoid of the F<sub>α</sub> type from fish.<sup>1</sup> Although this claim has recently been retracted,<sup>2</sup> the key issue regarding the occurrence of C<sub>22</sub>-prostanoids in marine organisms remains unresolved. It has been proposed that C<sub>22</sub>-prostanoids can be synthesized in mammalian tissue from 7,10,13,16-(Z)-docosatetraenoic (adrenic) acid.<sup>3</sup> On the other hand, studies in these laboratories have shown that DCHA is not a substrate for mammalian PG synthetase from ram seminal vesicles or human platelets and is in fact a potent competitive inhibitor of the conversion of arachidonate to PGs (ca. 10<sup>4</sup> times that of aspirin on a molar basis).<sup>4</sup> In order to demonstrate whether C<sub>22</sub>-prostanoids occur naturally and to evaluate their biological effects, it is clearly desirable to synthesize the compounds in question. We describe herein two processes for the synthesis of the E- and F<sub>α</sub>-type prostaglandins which would be derived from DCHA. These syntheses employ standard PG precursors<sup>5</sup> and illustrate two new methods for generating the required ω-appendage which are applicable to the synthesis of PG<sub>3s</sub> such as PGF<sub>3α</sub> and PGE<sub>3</sub>. Although the PG<sub>3s</sub> have been synthesized previously,<sup>6</sup> their preparation until now has been much more difficult than those for the PG<sub>1</sub> or PG<sub>2</sub> series.

The first process for the synthesis of ω-3 PGs utilized as a key reagent dimethyl (Z)-(2-oxohept-4-enyl)phosphonate (**1**). This new substance was synthesized in two steps as follows. Reaction of methyl sorbate with the lithio derivative of dimethyl methylphosphonate<sup>7</sup> in tetrahydrofuran (THF) at -78 °C for 8 h afforded after extractive isolation and column chromatography on silica gel (sg) 52% of dimethyl (E,E)-(2-oxohepta-3,5-dienyl)phosphonate (**2**) as a colorless liquid. Hydrogenation of freshly distilled **2** with 10 mol % of (methyl benzoate)chromium tricarbonyl<sup>8</sup> (750 psi H<sub>2</sub>, methylene chloride, 140-150 °C, 1.5 h)



proceeded by stereospecific 1,4-addition to give, after sg chromatography and distillation, **1**, bp 90 °C (0.01 mm), in 77% yield. Horner-Emmons coupling of **1** with optically active aldehyde **3**,<sup>6</sup> unsuccessful under a wide variety of conditions, proceeded as desired by using a solution of sodium methoxide (methanol free) in dry THF at -78 °C for 1 min to convert **1** to its anion, promptly adding **3**, raising the temperature to 0 °C rapidly, and allowing the reaction to proceed at 0 °C for 3 h. Quenching with acetic acid, extractive isolation, and chromatography on sg provided keto lactone **4** in 50% yield.<sup>9</sup> Cleavage of the tetrahydropyranyl group of **4** (pyridinium tosylate in methanol at 50 °C for 1.5 h) and carbonyl reduction using diisobutylaluminum 2,6-di-*tert*-butyl-4-methylphenoxide<sup>10</sup> at -78 °C in toluene afforded with 91:9 selectivity (85% yield) the required 15-*S*-diol **5** which readily separated from the less polar 15-*R* isomer by sg chromatography using 1:1 benzene-ethyl acetate for elution.<sup>11</sup> The minor 15-*R* isomer upon treatment with Attenburrow manganese dioxide in methylene chloride was converted to the corresponding 15-ketone and recycled.

Transformation of **5** into the bis(tetrahydropyranyl) ether of C<sub>22</sub>-PGF<sub>4α</sub> (**6**) was accomplished in 65% overall yield by the following sequence: (1) tetrahydropyranylation of **5** using 3 equiv of dihydropyran in methylene chloride with camphor-10-sulfonic acid catalyst at 0 °C for 20 min (99% yield); (2) lactone to lactol reduction using 1.6 equiv of diisobutylaluminum hydride in toluene at -78 °C for 30 min (99% yield); (3) Wittig reaction with the ylide from (Z)-7-(triphenylphosphonio)hept-4-enoate (generated using sodium bis(trimethylsilyl)amide in THF-toluene) at -40 °C then at 0 °C for 3.5 h; (4) esterification (CH<sub>2</sub>N<sub>2</sub>), sg chromatography and saponification.

The methyl ester of **6** was transformed into C<sub>22</sub>-PGF<sub>4α</sub> (**7**) by THP cleavage (pyridinium tosylate in methanol at 55 °C for 1 h) followed by saponification with 0.5 N sodium hydroxide in 50% aqueous methanol at 55 °C for 2.5 h (90% overall yield). Conversion of **6** to C<sub>22</sub>-PGE<sub>4</sub> (**8**) was accomplished by Jones oxidation at 0 °C for 30 min followed by THP cleavage (6:3:1 acetic acid-water-THF, 35 °C for 8 h).

A second route to **5** was developed which also started from lactone aldehyde **3** via the enal **9**, which was produced from **3** by the sequence: (1) RCHO → RCH=CHCOOC<sub>2</sub>H<sub>5</sub> chain extension using the sodium derivative of triethylphosphonoacetate

(1) Mai, J.; Goswami, S. K.; Bruckner, G.; Kinsella, J. E. *Prostaglandins* **1981**, *21*, 691.

(2) German, B.; Bruckner, G.; Kinsella, J. *Prostaglandins* **1983**, *26*, 207.

(3) Sprecher, H.; Van Rollins, M.; Sun, F.; Wyche, A.; Needleman, P. J. *Biol. Chem.* **1982**, *257*, 3912.

(4) Corey, E. J.; Shih, C.; Cashman, J. R. *Proc. Natl. Acad. Sci. U.S.A.* **1983**, *80*, 3581.

(5) Corey, E. J.; Schaaf, T. K.; Huber, W.; Koelliker, U.; Weinshenker, N. M. *J. Am. Chem. Soc.* **1970**, *92*, 397 and references cited therein.

(6) Corey, E. J.; Shirahama, H.; Yamamoto, H.; Terashima, S.; Venkateswarlu, A.; Schaaf, T. K. *J. Am. Chem. Soc.* **1971**, *93*, 1490.

(7) (a) Corey, E. J.; Kwiatkowski, G. T. *J. Am. Chem. Soc.* **1966**, *88*, 5654. (b) Corey, E. J.; Vlattas, I.; Andersen, N. H.; Harding, K. *Ibid.* **1968**, *90*, 3247.

(8) (a) Frankel, E. N.; Butterfield, R. O. *J. Org. Chem.* **1969**, *34*, 3930.

(b) Frankel, E. N.; Selke, E.; Glass, C. A. *J. Am. Chem. Soc.* **1968**, *90*, 2446.

(c) Mahaffy, C. A. L.; Pauson, P. L. *Inorg. Synth.* **1979**, *19*, 154.

(9) This yield has not been optimized with respect to reaction temperature and probably can be increased.

(10) Iguchi, S.; Nakai, H.; Hayashi, M.; Yamamoto, H.; Maruoka, K. *Bull. Chem. Soc.* **1981**, *54*, 3033.

(11) Thin-layer chromatographic (TLC) R<sub>f</sub> values (sg, 1:1 benzene-ethyl acetate) were 0.19 and 0.26 for **5** and the 15-*R* diastereomer, respectively.